

**Endo Agar**

For confirmation of the presumptive test for members of the coliform group from clinical and non-clinical samples.

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| REF:V.1/EN01.100 REF: V.1/EN01.500  | 100 Gram 500 Gram  | REF: V.1/EN01.250  | 250 Gram  |

**CLINICAL SIGNIFICANCE**

Endo Agar was developed by Endo to differentiate gram-negative bacteria on the basis of lactose fermentation, while inhibiting gram- positive bacteria (1). Inhibition of the later was achieved without the use of bile salts as was traditionally used. Endo was successful in inhibiting gram-positive bacteria on his medium by the incorporation of sodium sulphite and basic fuchsin. The resulting Endo Agar, also known as Fuchsin Sulphite and Infusion Agar, was used to isolate the typhoid bacilli. Many modifications of this media have been done over the years. Endo Agar is recommended by APHA as an important medium in the microbiological examination of water and wastewater, dairy products and foods (2, 3, and 4). Endo Agar is used to confirm the detection and enumeration of coliform bacteria following presumptive test of drinking water. It is also used for the detection and isolation of coliforms and faecal coliforms from milk, dairy products and food.

**METHOD PRINCIPLE**

The medium contains peptone which provide nitrogen, carbon, vitamins and minerals required for bacterial growth. Sodium sulphite and basic fuchsin make this medium selective by suppressing gram-positive organisms. Coliforms produce pink colonies on fermentation of lactose while lactose non-fermenters produce colorless colonies on the medium.

With Escherichia coli. This reaction is very pronounced as the fuchsin crystallizes, exhibiting a permanent greenish metallic luster (fuchsin luster) to the colonies. Medium should be stored away from light to avoid photo-oxidation

**MEDIA COMPOSITION**

|  |  |
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| Item  | Formula per liter of medium  |
| * Peptone
* Lactose
* Dipotassium hydrogen phosphate
* Sodium sulphite
* Basic fuchsin
* Agar
 | 10.00 gm 10.00 gm 3.500 gm 2.500 gm 0.500 gm 15.00 gm  |

**PRECAUTIONS AND WARNINGS**

Media to be handled by entitled and professionally educated person. Do not ingest or inhale.

Good Laboratories practices using appropriate precautions should be followed in:

* Wearing personnel protective equipment (overall, gloves, glasses,).
* Do not pipette by mouth.
* In case of contact with eyes or skin; rinse immediately with plenty of soap and water. In case of severe injuries; seek medical advice immediately.
* Respect country requirement for waste disposal.

S56: dispose of this material and its container at hazardous or special waste collection point.

S57: use appropriate c o n t a i n e r t o a v o i d e n v i r o n m e n t a l

Contamination.

S61: avoid release in environment.

For further information, refer to the Endo agar material safety data sheet.

STORAGE AND STABILITY

**Lab.Vie**. Endo Agar dehydrated media are stable until expiration date stated on label when properly stored 10-30°C. The prepared medium should be stored at 2-8°C. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to avoid lump development due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in a dry ventilated area protected from extremes of temperature and sources of ignition. Seal the container tightly after use. Product performance is best if used within stated expiry period.

Final pH 7.5 ± 0.2 at 25°C

MEDIA PREPARATION

Suspend 41.5 grams in 1000 ml purified/distilled water. Heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs. pressure (121°C) for 15 minutes. Mix well before pouring into sterile Petri plates. If the solidified culture medium is somewhat too red, then to remove the colour add a few drops (max. 1 ml/litre) of a freshly prepared 10% Sodium sulphite solution and boil.

Deterioration

The color of **Lab.Vie**. Endo Agar medium is Light pink to purple homogeneous free flowing powder. If there are any physical changes, discard the medium.

The hydrated medium is Orange pink coloured, clear to slightly opalescent gel with fine precipitate forms in Petri plates, media should not be used if there are any signs of deterioration (shrinking, cracking, or discoloration), and contaminations.

SPECIMEN COLLECTION AND PRESERVATION

Clinical samples - urine; Food and dairy samples; Water samples.

EQUIPMENT REQUIRED NOT PROVIDED

* Sterile cups
* Sterile petri-dishes
* Incubator

PERFORMANCE CHARACTERISTICS

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature

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| SYMBOLS IN PRODUCT LABELLING  |
| IVD  | For in-vitro diagnostic use Batch Code/Lot number   Catalogue Number Temperature Limitation Expiration Date Manufactured by  | Number of <n> test in the pack Caution  Do not use if package is damaged   Consult Instruction for use    |
| LOT |
|   |
| REF     |
|  | Test Organisms  | Growth  | Colour of Colony  |  |
| Klebsiella aerogenes ATCC 29212  | good- luxuriant  | pink  |
| Escherichia coli ATCC 25922  | good- luxuriant  | pink to rose red with metallic sheen  |
| Klebsiella pneumoniae ATCC 13883  | good- luxuriant  | pink, mucoid   |
|  Enterobacter cloacae ATCC 13047  | good  | pink  |  |
| Proteus vulgaris ATCC 13315  | good- luxuriant  | colourless to pale pink  |
| Pseudomonas aeruginosa ATCC 27853  | good- luxuriant  | colourless, irregular  |
| Salmonella Typhi ATCC 6539  | good- luxuriant  | colourless to pale pink  |
| Staphylococcus aureus subsp.aureus ATCC 25923  | inhibited  |   |
| Bacillus subtilis subsp. spizizenni ATCC 6633  | inhibited  |   |

QUALITY CONTROL

To ensure adequate quality control, it is recommended that positive and negative control included in each run. If control values are found outside the defined range, check the system performance. If control still out of range please contact **Lab.Vie**. technical support.

REFERENCES

1. Endo S., 1904, Zentralbl. Bakteriol., Abt. 1, Orig.35:109-11
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3. Salfinger Y., and Tortorello M.L. , 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.
4. Wehr H. M. and Frank J. H., 2004, Standard Methods for the Microbiological Examination of Dairy Products, 17th Ed.,APHA Inc., Washington, D.C.
5. Isenberg, H.D. Clinical Microbiology Procedures Handb0ook. 2nd Edition.
6. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W.(2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.

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